

Interview
THIS NEWSLETTER
FEATURES AN
INTERVIEW WITH
CHARLES GRIFFITHS

PAGE 2



BIBLIOGRAPHY
 PAGE 4

TAXON LIST
 PAGE 54

LETTER FROM
SIDOROV
 PAGE 63

SWISS AMPHIPODA
 PAGE 64

PATAGONIAN
AMPHIPODA
 PAGE 66

AMPHIPOD NEWSLETTER 43

Dear Amphipodologists,

AN 43 arrives only just after the 18th ICA in Dijon, where we again met many old friends and made many new ones. Sadly we have also in 2019 lost two dear colleagues. John Holsinger and Augusto Vigna Taglianti. An extensive in memoriam for John Holsinger has been written by David Culver (see bibliography), while we hope to be able to include an in memoriam for Augusto Vigna Taglianti (Roma) in AN 44. We have an interview with Charles Griffiths in Cape Town, the foremost author on S. African amphipods, but someone who never has made it to our conferences. There are also shorter contributions by Roman Alther and Dmitry Sidorov as well as a report from a workshop on Patagonian amphipoda. We are very happy to hear from you - and we are pleased to present both small and large reports from your work, workshops and gatherings! Information about getting in contact with the newsletter is at page 63.

*Statistics from
 this Newsletter*

53 new genera

72 new species

The bibliography this time contains 392 papers. The number of new species contained in it is with 72 maybe a bit less than in the earlier newsletters, but 53 new genera in one Newsletter is probably a record, due mainly to Jim Lowry & Alan Myers revisionary work on the Talitroidea. The higher classification of our beloved amphipods is clearly still in a state of flux.

We are very grateful to Tammy Horton for much help with AN43.

Wim and Anne Helene

Interview with Emeritus Prof Charles Griffiths, University of Cape Town

My career as an amphipodologist started completely by accident back in 1970, when I applied for a PhD scholarship to study larval biology of marine invertebrates at the University of Cape Town. The then Professor, the famous polychaete taxonomist John Day, informed me in the interview that he had already filled that post, but had another opportunity that he could offer me studying the taxonomy of amphipods. I had no idea what an amphipod was and zero taxonomic experience, but being too embarrassed to ask, agreed to accept the post! The next Monday I arrived for work and John Day took me into a large room next to his laboratory in which all the invertebrates collected during the past 30 years of his teams' ecological survey work were stored. 'Here is the collection' he told me 'your job is to identify all the amphipods and please don't bother me'. For the next 3 years I thus worked my way through the over 100 000 stored specimens, teaching myself the techniques from scratch, mostly using the then just published JL Barnard's 1969 'Families and Genera of Marine Gammaridean Amphipoda', which was my 'bible' and lifesaver! Of course the collections contained many new species and I was lucky to be able to publish these in a series of papers in Annals of the South African Museum, culminating in a guide book to the regional amphipod fauna, which today remains the only such resource.

After my PhD and a short stint as a research officer I was appointed to the academic staff at UCT, where I have continued to work for nearly 50 years. Having spend my PhD mostly staring down a microscope I was initially not keen to continue with taxonomic work, but as I was now the only regional



Charles and his wife Roberta kayaking among the kelp where his newest amphipod-species, named after his kelp-researcher wife are found.

amphipod taxonomist, other researchers continued to send me interesting material for identification, so along with my main ecological research (mostly on kelp beds, mussels and predator-prey interactions) I continued to publish the odd species descriptions. Later on in my career I became involved as leader of the African group of Census of Marine

Life Programme and returned to doing more taxonomic work, although mostly on groups other than amphipods. Together with various students and co-authors I have, for example, documented many additions to the regional fauna of anemones, barnacles, mysids, decapod crustacea, sea-urchins, brittlestars, ascidians and even one new species of fish). Overall, I am now an author of papers describing over 100 species new to science and adding more than 200 others to the South African fauna. My recent passion has also been the production of popular photographic field guides and I am an author of the standard field guides to South African Marine Life, to The Insects and to Freshwater Life.

My most memorable amphipod moments both involve the discovery of species that were subsequently named after family members. The first of these involved the discovery of a new genus of freshwater paramelitid amphipod nearly 30 years ago. While driving up a narrow mountain pass on a family holiday, my son Matthew, then about 3, urgently needed a toilet, so I pulled into a narrow verge where a small stream crossed the road. While he was busy I turned some rocks over in the stream and was surprised to find amphipods, which we not known from that part of South Africa. These were later described as the types of a new genus *Mathamelita*, in honour of Matthews, whose bodily needs led to their discovery. Another more recent experience was the discovery of a large kelp-boring amphipod, which I named after my wife Roberta, as we met back in the 1970s when working together as kelp bed researchers.



All four members of the Griffiths-family holding a photo of the species that have been named after them. Daughter Melinda (nudibranch *Leminda millecra* R. Griffiths, 1985), son Matthew (amphipod *Mathamelita aequicaudata* Stewart & C. Griffiths, 1995), Charles (amphipod *Griffithsius latipes* Griffiths, 1976 - (*Griffithsius* Jarrett & Bousfield, 1994)) and wife Roberta (amphipod *Sunamphitoe roberta* C. Griffiths, 2019).

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[dx.doi.org/10.11646/zootaxa.4568.1.4](https://doi.org/10.11646/zootaxa.4568.1.4) (*Dorotea papuensis* n. gen., n. sp. (N. of Laughlan archipelago, Solomon Sea, PNG). The species *Eusiroides aberrantis* is transferred to *Dorotea*. A key to eusirid genera is provided.)

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Just, J. 2019. Siphonoecetini Just, 1983 (Crustacea, Amphipoda, Ischyroceridae) 13: Western Australian species of Bubocorophiina in *Rhinoecetes*, *Cephaloecetes*, *Sinoecetes*, *Borneoecetes* and *Pararhinoecetes* gen. nov.. ---- *Zootaxa* 4554, 101-140. <https://doi.org/10.11646/zootaxa.4554.1.3> (Deals with *Rhinoecetes sinuduopopulus* n. sp. (Two Peoples Bay), *Rh. rockinghamia* n. sp. (Rockingham), *Rh. makritricjoma* n. sp. (Rottnest Island), *Rh. lowryi* n. sp. ((Rottnest Island), *Rh. caetus* n. sp. (Shark Bay), *Rh. karkharius* n. sp. (Shark Bay), *Rh. wamus* n. sp. (Cape Range NP), *Rh. setosus* n. sp. (Cape Range NP), *Borneoecetes minimus* n. sp. (off Port Hedland), *Sinoecetes reni* n. sp. (King George Sound) and *Pararhinoecetes bicornis* n. gen., n. sp. (North West Shelf). A key to all Western Australian Bubocorophiini is included.)

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Michaud, K. M., K. A. Emery, J. E. Dugan, D. M. Hubbard & R. J. Miller 2019. Wrack resources use by intertidal consumers on sandy beaches. ---- *Estuarine, Coastal and Shelf Science* 221, 66-71. <https://doi.org/10.1016/j.ecss.2019.03.014> (*Megalorchestia* spp.)

Mijošek, T., V. Filipović Marijić, Z. Dragun, D. Ivanković, N. Krasnići, M. Erk, S. Gottstein, J. Lajtner, M. Serić Perić & R. Motoničkin Kepčija 2019. Comparison of electrochemically determined metallothionein concentrations in wild freshwater salmon fish and gammarids and their relation to total and cytosolic metal levels. ---- *Ecological Indicators* 105, 188-198. <https://doi.org/10.1016/j.ecolind.2019.05.069> (*Gammarus balcanicus* and *Echinogammarus acarinatus*)

Mikhaylova, T. A., D. A. Aristov, A. D. Naumov, S. S. Malavenda, O. N. Savchenk & K. L. Bijagov 2019. Diversity and structure of epibenthic communities of the red algae zone in the White Sea. ---- *Polar Biology* 42, 953-968. <https://doi.org/10.1007/s00300-019-02488-2>

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Moraes, M. L. L. de 2018 (*Temporal variation of vagile macrofauna-specially Amphipoda—associated with the Sargassum furcatum of the São Sebastião Island.*) ---- PhD Thesis, University of Campinas (Not seen)

Morales-Núñez, A. G. & P. Chignu 2019. Abundance, distribution, and species composition of amphipods associated with macroalgae from shallow waters of the Maryland Coastal Bays, USA. ---- *Marine Biodiversity* 49, 175-191. <https://doi.org/10.1007/s12526-017-0779-z> .

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doi.org/10.1016/j.rsma.2019.100685 (Amphipods, primarily Ampithoidae and Dogielinotidae, identified and studied molecularly, but not named.)

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Myers, A. A., J. K. Lowry & R. S. K. Barnes 2018. First record of the genus *Eriopisella* Chevreux, 1920 (Crustacea, Amphipoda, Senticaudata, Eriopisidae) from Australia, with the description of a new species, *Eriopisella moretoni* sp. nov. ---- *Zootaxa* 4514, 256-262. <http://dx.doi.org/10.11646/zootaxa.4514.2.8> (*E. moretoni* n. sp. from Moreton Bay, Queensland. With a key to world *Eriopisella* spp.)

Myers, A. A., R. A. Sreepada & S. V. Sanaye 2019. A new species of *Grandidierella* Coutière, 1904, *G. nioensis* sp. nov. (Amphipoda, Aoridae), from the east coast of India. ---- *Zootaxa* 4541, 119-124. <http://dx.doi.org/10.11646/zootaxa.4544.1.7> (From Machilipatnam, Andhra Pradesh. With a key to the *G. mahafalensis* species complex).

Nakamura, Y., T. Nakano, Y. Ota & K. Tomikawa 2019. A new species of the genus *Elasmopus* from Miyako Island, Japan (Crustacea, Amphipoda, Maeridae). ---- *Zootaxa* 4544, 395-406. <https://doi.org/10.11646/zootaxa.4544.3.5> (*E. nkjaf* n. sp.; Miyako Island is in the Ryu Kyu Islands. A key to all *Elasmopus* in Japanese waters is provided.)

Nakano, T. 2018. Mandatory changes of specific names to agree in gender with *Talitriator* Methuen, 1913, which is masculine (Crustacea: Amphipoda: Talitridae). ---- *Zootaxa* 4483: 188-190. <http://www.mapress.com/j/zt/article/view/zootaxa.4483.1.8/16060> (*Talitriator setosa* and *T. calva* should change to *T. setosus* and *T. calvus* respectively)

Nakano, T. & K. Tomikawa 2018. Reassessment of the groundwater amphipod *Paramoera relict* synonymiizes the genus *Relictomoera* with *Paramoera* (Crustacea: Amphipoda: Pontogeneiidae). ---- *Zoological Science* 35, 459-467. <https://doi.org/10.2018/zs180058> (*P. relict* fully redescribed and molecularly characterized. The genus *Relictomoera* was based on an erroneous observation.)

Nakano, T., K. Tomikawa & M. J. Grygier 2018. Rediscovered syntypes of *Procrangonyx japonicus*, with nomenclatural consideration of some crangonyctoidean subterranean amphipods (Crustacea: Amphipoda: Allocrangonyctidae, Niphargidae, Pseudocrangonyctidae). ---- *Zootaxa* 4532, 86-94. <http://dx.doi.org/10.11646/zootaxa.4532.1.4> (*Procrangonyx* has been validly described and *Eocrangonyx* is an objective junior synonym, based on the same type species. The date of publication of some Schellenberg papers is discussed.)

Ng, C. S. L., K. B. Toh, T. C. Toh, J. Y. Ng, P. R. Cheo, K. Tun & L. M. Chou 2019. Distribution of soft bottom macrobenthic communities in tropical marinas of Singapore. ---- *Urban Ecosystems* 22, 443-453. <https://doi.org/10.1007/s11252-019-0828-4> .

Ólafsdóttir, J. H., J. G. Þorbjörnsson, B. J. Kristjánsson & J. S. Ólafsson 2019. Invertebrate biodiversity in cold groundwater fissures in Iceland. ---- *Ecology and Evolution* 9(11), 6399-6409 <https://doi.org/10.1002/ece3.5213> (No amphipods)

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Ótero-Ferrer, F., E. Mannardi, M. Cosme, A. Falace, J. A. Montiel-Nelson, F. Espino, R. Haroun & F. Tuya 2019. Early-faunal colonization patterns of discrete habitat units: A case study with rhodolith-associated vagile macrofauna. ---- *Estuarine, Coastal and Shelf Studies* 218, 9-22. <https://doi.org/10.1016/j.ecss.2018.11.020>

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Ozga, A. V., V. da S de Castro & D. de S. Castiglione 2018. Population structure of two freshwater amphipods (Crustacea: Peracarida: Hyalellidae) from southern Brazil. ---- *Nauplius* 26, e 2018025 <http://dx.doi.org/10.1590/2358-2936e2018025> (*H. georginae* and *H. gauchensis*)

Pacioglu, O., N. Ianovici, M. N. Filimon, A. Sinitean, G. Iacob, H. Barabas, A. Pahomi, A. Acs, H. Muntean & L. Pârvulescu 2019. The multifaceted effects induced by floods on the macroinvertebrate communities inhabiting a sinking cave stream. ---- *International Journal of Speleology* 48, 167-177. <https://doi.org/10.5038/1827-806X.48.2.2239> .

Palatov, D. M. & A. M. Sokolova 2019. Stygobiotic faunal elements in spring assemblages of West Transcaucasia. ---- *Ecosystem Trasformation* 2, 35-43. (Not seen) http://www.ecosysttrans.com/publikatsii/detail_page.php?ID=95

Pandey, V. & G. Thiruchitrambalam 2019. Spatial and temporal variability of sandy intertidal microbenthic communities and their relationship with environmental factors in a tropical island. ---- *Estuarine, Coastal and Shelf Science* 224, 73-83. <https://doi.org/10.1016/j.ecss.2019.04.045>
(A study from the Andaman Islands)

Pařil, P., C. Leigh, M. Polášek, R. Sarremejane, P. Řezníčková, A. Dostálová & R. Stubbington 2019. Short-term streambed drying events alter amphipod population structure in a central European stream. ---- *Fundamental and Applied Limnology*, in press. <https://doi.org/10.1127/fal/2019/1164> (*Gammarus fossarum*)

Parry, R. & S. Asgari 2019. Discovery of novel crustacean and cephalopod flaviviruses: insights into evolution and circulation of flaviviruses between marine invertebrates and vertebrate hosts. ---- *Journal of Virology*, in press <https://doi.org/10.1128/JVI.00432-19> (i.a. from *Gammarus chevreuxi* and *G. pulex*.)

Patel, T., H. Robert, C d'Udekem d'Acoz, K. Martens, I. de Mesel, S. Degraer & I. Schön 2018. Biogeography and community structure of abyssal scavenging Amphipoda (Crustacea) in the Pacific Ocean. ---- *Biogeosciences Discussion*, in review. <https://doi.org/10.5194/bg-2018-347> .

Pauli, N.-C., F. Paiva & E. Briski 2018. Are Ponto-Caspian species able to cross salinity barriers? A case study of the gammarid *Pontogammarus maeoticus*. ---- *Ecology and Evolution* 8(19), 9817-9826. <https://doi.org/10.1002/ece3.4461> (Yes to lower salinity, no to higher salinity - for this species...)

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Paz-Rios, C. E., N. Simões & D. Pech 2019. Species richness and spatial distribution of benthic amphipods (Crustacea: Peracarida) in the Alacranes Reef National Park, Gulf of Mexico. ---- *Marine Biodiversity* 49, 673-682. <https://doi.org/10.1007/s12526-017-0843-8> .

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Perez Schultheiss, J., L. Fernandez Parra & K. Ayala 2018. (Taxonomic revision of the genus *Orchestoidea* Nicolet, 1849 (Crustacea: Amphipoda: Talitridae).) ---- pp 79-96 in Informe del Fondo de Apoyo para la Investigación Patrimonial (Faip) (In Spanish. Two new species are described , but not named)

Perrot-Minot, M.-J., A. Chaumot, G. Caillot, H. Quéau, N. Delorme & O. Geffard 2018. Combined effects of parasitism and anthropogenic stressors in the freshwater amphipod *Gammarus fossarum*: Impacts on multiple traits. ---- P. 360 in In: Update on selected topics in acanthocephalan parasites research. *Helminthologia* 55, 350-362.
<https://doi.org/10.2478/helm-2018-0023>

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Pezy, J.-P., C. Delecruin, A. Baffreau, O. Basuyaux & J.-C. Dauvin 2019. Anthropogenic impact of oyster farming on macrofauna biodiversity in an eelgrass (*Zostera marina*) ecosystem of the English Channel. ---- *Ecological Indicators* 106, 105480. <https://doi.org/10.1016/j.ecolind.2019.105480> .

Picone, M., M. Bergamin, E. Delaney & A. Volpi Ghirandini 2018. Assessment of whole-sediment chronic toxicity using sub-lethal endpoints with *Monocorophium insidiosum*. ---- *Ecotoxicology* 27(9), 1237-1248. <https://doi.org/10.1007/s10646-018-1977-6> .

Piertney, S. B. & A. Jamieson 2018. *Genetic structure within and between deep-ocean trenches in the hadal amphipod Bathycallisoma*. ---- P. 33 in 50th Population genetics Group Meeting 2017. (*B. schellenbergi*)

Podlesińska, W. & H. Dąbrowska 2018. Amphipods in estuarine and marine quality assessment—a review. ---- *Oceanologia* 61, 179-196. <https://doi.org/10.1016/j.oceano.2018.09.002>

Polgar, J., D. Zeballos, J. Vargas, M. Aldana, P. Manriquez, K. Manriquez, P. A. Quijon, S. Widdicombe, C. Anguita, D. Quintanilla & C. Duarte 2018. Endogenous cycles, activity patterns and energy expenditure of the intertidal fish is modified by Artificial Light Pollution at Night (ALAN). ---- *Environmental Pollution* 244, 361-366. <https://doi.org/10.1016/j.envpol.2018.10.063>

Poore, A. G. B., L. Gutow, A.-N. Lörz & M. Thiel 2018. Nest building by a small mesograzer limits blade size of the giant kelp *Macrocystis pyrifera*. ---- *Marine Biology* 165: 184 <https://doi.org/10.1007/s00227-018-3444-6> (*Pseudopleonexes lessoniae*)

Power, C., J. Balli-Garza, D. Evans, B. F. Nowak, A. R. Bridle & N. J. Bott 2019. Detection of *Miamiensis avidus* (Ciliophora: Scuticiliatia) and *Cardicola* spp. (Trematoda: Aporocotylidae) DNA in biofouling from Southern Bluefin Tuna, *Thunnus maccoyii* pontoons off Port Lincoln, South Australia. ---- *Aquaculture* 502, 128-133. <https://doi.org/10.1016/j.aquaculture.2018.12.027>

Poynton, H., C. Chen, S. L. Alexander, K. M. Major, B. J. Blalock & J. Unrine 2019. Enhanced toxicity of environmentally transformed ZnO nanoparticles relative to Zn ions in the epibenthic amphipod *Hyaella azteca*. ---- *Environmental Science Nano* 6, 325-340. <https://doi.org/10.1039/C8EN00755A> .

Premate, E., T. Volk, D. Copilas-Cioceanu, Z. Fiser, A. Jemc Kokaj, T. Delic & C. Fiser 2018. Locomotion of *Niphargus* amphipods from cave lakes and streams. ---- *ARPHA Conference Abstracts* 1, e30389. <https://doi.org/10.3897/aca.1.e30389>

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Puzin, C. & J. Pétilon 2019. Contrasted responses of dominant ground-dwelling arthropods to landscape salt marsh fragmentation. ---- *Estuarine, Coastal and Shelf Science* 224, 138-141. <https://doi.org/10.1016/j.ecss.2019.05.002> (i.a. *Orchestia gammarellus*)

Quiles, A., K. Bacela- Spychalska, M. Teixeira, N. Lambin, M. Grabowski, T. Rigaud & R. A. Wattier 2019. Microsporidian infections in the species complex *Gammarus roeselii* (Amphipoda) over its geographical range: evidence for both host-parasite co-diversification and recent host shifts. ---- *Parasites & Vectors* 12: 327 <https://doi.org/10.1186/s13071-019-3571-z>

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obuchii, *L. ouraensis*, *L. togatta*, *L. taribe*, *L. trulla*, *L. batillum* n. sp. (Dongsha Atoll), *L. cracentis* n. sp. (Dongsha Atoll) and *Paranamixis lunata* n. sp. (Dongsha Atoll))

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Polymorphus minutus. ---- P. 358 in In: Update on selected topics in acanthocephalan parasites research. *Helminthologia* 55, 350-362. <https://doi.org/10.2478/helm-2018-0023>

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Compilation of Amphipod relevant literature

Please tell the AN editors and Olli Coleman about your recent publications on amphipods - and send a pdf of your paper. Olli can include it on the server and the editors can include it in the bibliography....

NEW TAXA

GENERA

<i>Albidiator</i> Lowry & Myers, 2019	Talitridae
<i>Allorchestoides</i> Wongkamhaeng, Dumrongrojwattana & Shin, 2018	Dogielinotidae
<i>Amphiatlantica</i> Lowry & Myers, 2019	Talitridae
<i>Asiaorchestia</i> Lowry & Myers, 2019	Talitridae
<i>Bulychevia</i> Lowry & Myers, 2019	Talitridae
<i>Calviator</i> Lowry & Myers, 2019	Talitridae
<i>Canariorchestia</i> Lowry & Myers, 2019	Talitridae
<i>Chaetoniphargus</i> Karaman & Sket, 2019	Niphargidae
<i>Chevreurxiana</i> Lowry & Myers, 2019	Talitridae
<i>Chevreurxiopsis</i> Halfter & Coleman, 2019	Thoriellidae
<i>Cognateosymtes</i> Labay, 2018	Pleustidae
<i>Dallwitzia</i> Lowry & Myers, 2019	Talitridae

Defeo Lowry & Myers, 2019	Talitridae
Dendrorchestia Lowry & Myers, 2019	Talitridae
Deshurleyella Lowry, Myers & Nakano, 2019	Talitridae
Derzhavinia Lowry & Myers, 2019	Talitridae
Dinarogammarus Sket & Hou, 2018	Gammaridae
Dorotea Corbari, Frutos & Sorbe, 2019	Eusiridae
Dracorchestia Lowry & Myers, 2019	Talitridae
Fleuria Lowry & Myers, 2019 (NB homonym)	Talitridae
Fleuriella Lowry, Myers & Nakano, 2019	Talitridae
Galaporchestia Lowry & Myers, 2019	Talitridae
Hermaniator Lowry & Myers, 2019	Talitridae
Houlia Lowry & Myers, 2019	Talitridae
Hurleyella Lowry & Myers, 2019 (Homonym)	Talitridae
Iberogammarus Sket & Hou, 2018	Gammaridae
Ignamborchestia Lowry & Myers, 2019	Talitridae
Indiorchestia Lowry & Myers, 2019	Talitridae
Insulariator Lowry & Myers, 2019	Talitridae
Kaalorchestia Lowry & Myers, 2019	Talitridae
Kellyduncania Lowry & Myers, 2019	Talitridae
Kohuroa Lowry, Myers & Nakano, 2019	Talitridae
Laniporchestia Lowry & Myers, 2019	Talitridae
Laurenia Lowry & Myers, 2019	Talitridae
Leslieorchestia Lowry & Myers, 2019	Talitridae
Lutruwitiator Lowry & Myers, 2019	Talitridae
Mexitroides Lindeman, 1990 (upgraded)	Talitridae
Morinoia Lowry & Myers, 2019	Talitridae
Oamaru Lowry & Myers, 2019	Talitridae
Omaiorchestia Lowry & Myers, 2019	Talitridae
Opunorchestia Lowry & Myers, 2019	Talitridae
Orientogidiella Sidorov, Ranga Reddy & Shaik, 2018	Austroniphargidae
Pararhinoecetes Just, 2019	Siphonoecetinae
Pickorchestia Lowry & Myers, 2019	Talitridae
Propejanice Myers & Desiderato, 2019	Aoridae
Richardsoniella Lowry, Myers & Nakano, 2019	Talitridae
Selvacaprella Guerra-Garcia, Tato & Moreira, 2018	Caprellidae

<i>Sinbadorchestia</i> Lowry & Myers, 2019	Talitridae
<i>Snaresorchestia</i> Lowry & Myers, 2019	Talitridae
<i>Speziorchestia</i> Lowry & Myers, 2019	Talitridae
<i>Swaziator</i> Lowry & Myers, 2019	Talitridae
<i>Tasmanella</i> Lowry & Myers, 2019 (homonym)	Talitridae
<i>Wairua</i> Lowry & Myers, 2019 (homonym)	Talitridae

SPECIES

<i>aequanime</i> Gasca, 2018 (In Gasca & Browne 2018) (<i>Megalanceoloides</i>)	Megalanceoloidae
<i>anneheleneae</i> Fuchs, Coleman & Lörz, 2019 (<i>Syrrhoe</i>)	Synopiidae
<i>ariakensis</i> Ariyama, 2019 (<i>Austromaera</i>)	Maeridae
<i>batillum</i> White & Machida, 2018 (<i>Leucothoe</i>)	Leucothoidae
<i>bjarnii</i> Bellan-Santini, Kaim-Malka & Dauvin, 2018 (<i>Haploops</i>)	Ampeliscidae
<i>bicornis</i> Just, 2019 (<i>Pararhinoecetes</i>)	Siphonoecetinae
<i>caetus</i> Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
<i>carthaginiensis</i> Ayati & Piscart in Ayati et al., 2018 (<i>Echinogammarus</i>)	Gammaridae
<i>cebuensis</i> Just, 2018 (<i>Sebadexius</i>)	Dexaminidae
<i>corallina</i> Zettler, Freiwald & Guerra-Garcia, 2018 (<i>Aeginella</i>)	Caprellidae
<i>cracentis</i> White & Machida, 2018 (<i>Leucothoe</i>)	Leucothoidae
<i>djemoi</i> Dole-Olivier, Hafid & Piscart, 2018 (<i>Pseudoniphargus</i>)	Pseudoniphargidae
<i>doughertyensis</i> Cannizzaro & Sawicki in Cannizzaro et al., 2019 (<i>Stygobromus</i>)	Crangonyctidae
<i>egmao</i> Özbek & Günoglu, 2019 (<i>Gammarus</i>)	Gammaridae
<i>elvirae</i> Paz-Rios & Pech, 2019 (<i>Gammaropsis</i>)	Photidae
<i>franki</i> Halfter & Coleman, 2019 (<i>Chevreuxiopsis</i>)	Thoriellidae
<i>gegi</i> Marin, 2019 (<i>Niphargus</i>)	Niphargidae
<i>gotoensis</i> Ariyama, 2019 (<i>Quadrimaera</i>)	Maeridae
<i>hakuhoae</i> Takeuchi, Kihara & Matsumoto, 2019 (<i>Caprella</i>)	Caprellidae
<i>hamaticornis</i> Copilaş-Ciocianu, Zimţa & Petrusek, 2018 (<i>Gammarus</i>)	Gammaridae
<i>hawkingi</i> Jazdzewska & Ziemkiewicz, 2019 (<i>Bathyceradocus</i>)	Maeridae
<i>hindustanica</i> Sidorov, Ranga Reddy & Shaik, 2018 (<i>Bogidiella</i>)	Bogidiellidae

hinojosai Hughes & Lörz, 2019 (<i>Bircenna</i>)	Eophliantidae
iemanja Silvany & Senna, 2019 (<i>Colomastix</i>)	Colomastigidae
irenae Do Nascimento & Serejo, 2018 (<i>Puelche</i>)	Phoxocephalopsidae
jazdzewskii Rudolph, Coleman, Mamosz & Grabowski, 2018 (<i>Gammarus</i>)	Gammaridae
jimenoi Guerra-Garcia, Tato & Moreira, 2018 (<i>Selvacaprella</i>)	Caprellidae
karkharius Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
lagamarensis Myers & Desiderato, 2019 (<i>Propejanice</i>)	Aoridae
laleyei Gnohossou & Piscart, 2019 (<i>Quadrivisio</i>)	Maeridae
longidactylus Do Nascimento & Serejo, 2018 (<i>Puelche</i>)	Phoxocephalopsidae
lorestanensis Esmaeli-Rineh, 2018 (<i>Niphargus</i>)	Niphargidae
lubuskensis Karaman & Sket, 2019 (<i>Chaetoniphargus</i>)	Niphargidae
lowryi Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
lunata White & Machida (<i>Paranamixis</i>)	Leucothoidae
makritrichoma Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
marielle Silvany & Senna, 2019 (<i>Colomastix</i>)	Colomastigidae
manubrium Cannizzaro & Sawicki, In Cannizzaro et al., 2019 (<i>Crangonyx</i>)	Crangonyctidae
mazatlanensis Alarcon-Ortega & Caballo, 2019 (<i>Deutella</i>)	Caprellidae
minimus Just, 2019 (<i>Borneoecetes</i>)	Siphonoecetinae
miramirandella Alves, Neves & Johnsson, 2018 (<i>Quadrimaera</i>)	Maeridae
moretoni Myers, Lowry & Barnes, 2018 (<i>Eriopisella</i>)	Eriopisidae
mourae Do Nascimento & Serejo, 2018 (<i>Puelche</i>)	Phoxocephalopsidae
nioensis Myers, Sreepada & Sanaye, 2019 (<i>Grandidierella</i>)	Aoridae
nkjaf Nakamura, Nakano, Ota & Tomikawa, 2019 (<i>Elasmopus</i>)	Maeridae
pingxiangensis Zheng, Hou & Li, 2018 (<i>Bogidiella</i>)	Bogidiellidae
papuana Corbari, Frutos & Sorbe, 2019 (<i>Dorotea</i>)	Eusiridae
quebecois Bellan-Santini, Kaim-Malka & Dauvin, 2018 (<i>Haploops</i>)	Ampeliscidae
reducta Sidorov, Ranga Reddy & Shaik, 2018 (<i>Orientogidiella</i>)	Austroniphargidae
reni Just, 2019 (<i>Sinoecetes</i>)	Siphonoecetinae
roberta Griffiths, 2019 (<i>Sunamphitoe</i>)	Ampithoidae
rockinghamia Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
rosea Wongkamhaeng, Dumrongrojwattana & Shin, 2018 (<i>Allorchestoides</i>)	Dogielinotidae
ruffoi Do Nascimneto & Serejo, 2018 (<i>Phoxocephalopsis</i>)	Phoxocephalopsidae

sakhalinensis Labay, 2019 (<i>Cryptodius</i>)	Ochlesidae
seisuiiae Kodama & Kawamura, 2019 (<i>Bemlos</i>)	Aoridae
serraticoxae Labay, 2018 (<i>Cognateosymtes</i>)	Pleustidae
setibasis Ariyama, 2019 (<i>Quadrimeaera</i>)	Maeridae
setosus Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
sinuduopopulus Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
subterraneus Sidorov, Ranga Reddy & Shaik, 2018 (<i>Indoniphargus</i>)	Austroniphargidae
thieli Hughes & Lörz, 2019 (<i>Bircenna</i>)	Eophliantidae
trispinosa Silvany, Alves & Senna, 2019 (<i>Colomastix</i>)	Colomastigidae
tubulosa Silvany & Senna, 2019 (<i>Colomastix</i>)	Colomastigidae
tunetanus Ayati & Piscart, in Ayati et al. 2018 (<i>Echinogammarus</i>)	Gammaridae
ungulatus Momtazi, Maghsoudlou & Just, 2018 (<i>Cephaloecetes</i>)	Siphoecetini
vaderi Alves, Johnsson & Senna, 2019 (<i>Ceradocus</i> (<i>Denticeradocus</i>))	Maeridae
vitucoi Guerra-Garcia, Tato & Moreira, 2018 (<i>Liropus</i>)	Caprellidae
wakulla Drumm & Knight-Gray, 2019 (<i>Hyaella</i>)	Hyaellidae
wamus Just, 2019 (<i>Rhinoecetes</i>)	Siphonoecetinae
willyi Guerra-Garcia, Tato & Moreira, 2018 (<i>Liropus</i>)	Caprellidae
yemanjae Alves, Neves & Johnsson, 2018 (<i>Quadrimeaera</i>)	Maeridae

Taxonomic overview

Ampeliscidae

Haploops **bjarnii**, **quebecois**

Ampithoidae

Sunamphitoe **roberta**

Aoridae

Bemlos **seisuiiae**

Grandidierella **nioensis**

Propejanice lagamarensis

Austrophargidae

Indoniphargus **subterraneus**

Bogidiellidae

Bogidiella **hindustanica**, **pingxiangensis**

Orientogidiella reducta

Caprellidae

Aeginella **corallina**

Caprella **hakuhoae**

Deutella **mazatlanensis**

Liropus **vitucui**, **willyi**

Selvacaprella jimeno

Colomastigidae

Colomastix **iemanja**, **marielle**, **trispinosa**, **tubulosa**

Crangonyctidae

Crangonyx **manubrium**

Stygocromus **doughtertyensis**

Dexaminidae

Sebadexius **cebuensis**

Dogielinotidae

Allorchestoides rosea

Eophliantidae

Bircenna **hinojosai**, **thieli**

Eriopisidae

Eriopisella **moretoni**

Eusiridae

Dorotea papuana

Gammaridae

DinarogammarusEchinogammarus **carthaginiensis**, **tunetanus**Gammarus **egmao**, **hamaticornis**, **jazdzewskii****Iberogammarus**

Hyalellidae

Hyalella **wakulla**

Ischyroceridae Siphonoecetini

Cephaloecetes **ungulatus**

Leucothoidae

Leucothoe **batillum**, **cracentis**Paranamixis **lunata**

Maeridae

Austromaera **ariakensis**Bathyceradocus **hawkingi**Ceradocus **vaderi**Elasmopus **nkjaf**Quadrimaera **gotoensis**, **miramirandella**, **setibasis**, **yemanjae**Quadrivisio **laleyei**

Megalanceoloidae

Megalanceoloides **aequanime**

Niphargidae

Chaetoniphargus lubuskensisNiphargus **gegi**, **lorestanensis**

Ochlesidae

Cryptodius **sakhalinensis**

Photidae

Gammaropsis **elvirae**

Phoxocephalopsidae

Phoxocephalopsis **ruffoi**

Puelche **irenae, longidactylus, mourae**

Pleustidae

Cognateosymtes serraticoxae

Pseudoniphargidae

Pseudoniphargus **djemoi**

Siphonoecetinae

Borneoecetes **minimus**

Pararhinoecetes bicornis

Rhinoecetes **caetus, karkharius, lowryi, makritrichoma, rockinghamia, setosus, sinuduopopulus, wamus**

Sinoecetes **reni**

Synopiidae

Syrrhoe **anneheleneae**

Talitridae

Albidiator

Amphiatlantica

Asiaorchestia

Bulychevia

Calviator

Canariorchestia

Chevreuxiana

Dallwitzia

Defeo

Dendrorchestia

Derzhavinia

Deshurleyella

Dracorchestia

Fleuria

Fleuriella

Galaporchestia
Hermaniator
Houlia
Hurleyella
Ignamborchestia
Indiorchestia
Insulariator
Kaalorchestia
Kellyduncania
Kohuroa
Laniporchestia
Laurenia
Leslieorchestia
Lutruwitiator
Mexitroides (upgr.)
Morinoia
Oamaru
Omaiorchestia
Opunorchestia
Pickorchestia
Richardsoniella
Sinbadorchestia
Snaresorchestia
Speziorchestia
Swaziator
Tasmanella
Wairua

Thoriellidae

Chevreuxiopsis franki

Who helps Sidorov?



My name is Dmitry Sidorov. Currently, I'm employee of the East Asia Terrestrial Biodiversity Center of the Russian Academy of Sciences in Vladivostok and engaged in research mainly of amphipods and slightly isopods. The most interesting, but also a difficult object in terms of research is a minute crustaceans which are still a lot of unspecified in the Far Eastern region of Russia. At the moment I dispose of a decommissioned microscope Carl Zeiss NU-2 in a deplorable technical condition, on which I working for many years. Prior to this, this microscope served in the walls of the POI FEB RAS and was made in the ancient soviet times. It was a very good microscope, but already it has

anti-reflection coating covered with unremovable divorces, the light is darkened, there no mercury lighting lamps and collimating procedure is needed. I happened to try different ways to fix it, but did not achieve any tangible success. Due to the lack of the good financial opportunity to purchase a microscope as a part of a grant, I unfortunately were left without equipment. I would be grateful if someone could help in this situation. I need some kind of microscope with magnification up to 800X-1000X supplied with a drawing device (camera lucida), phase contrast is desirable (it can be amortized second-hand microscope). Perhaps I will be able to pay the shipping and/or customs duty. In gratitude, I will determine a collections of your amphipods, if you need it for your project. Or I will gather some living creatures from the vicinity of Vladivostok/Primorye for your research.

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Swiss amphipoda

In September, the first monograph on amphipods of Switzerland will be released. The Fauna Helvetica book #32 – Amphipoda will be published by the Swiss Center for the Cartography of the fauna (CSCF) and the Swiss Entomological Society (SEG). Whereas the monograph is written in German, the identification key is trilingual (DE, FR, EN). The 389 pages contain a comprehensive overview about the amphipods of Switzerland, their ecology and their faunistics. The species identification key is suitable for beginners and experts, and covers all species of Switzerland and adjacent biogeographic regions, including all *Niphargus* species. The detailed texts for all 40 species hitherto reported from Switzerland are illustrated with 125 drawings and color photographs, and distribution maps of Switzerland. Many more additional drawings are contained in the identification key. All drawings were specifically prepared for this book. Additionally, the book covers a few species that are hitherto not reported from Switzerland but to be expected in the near future. This, as well as the illustrated identification key, renders the book valuable not only for scientists in Switzerland but also for amphipodologists from adjacent regions. We, Florian Altermatt, Roman Alther, Cene Fišer, and Vid Švara, hope that the monograph will meet with wide interest. It may be ordered through the publisher at <https://cscf.abacuscity.ch/de/chf/1~1~Coll/Fauna-Helvetica> as soon as the book is released. Preordering is possible at <http://www.cscf.ch/cscf/de/home/publikationen.html> via the PDF document.

FAUNA HELVETICA
AMPHIPODA

SEG CSCF



Florian Altermatt
Roman Alther
Cene Fišer
Vid Švara

Inhalt

Einleitung	4
Dank	6
Summary	8
Allgemeiner Teil	11
Morphologie • Biologie und Verhalten • Ökologie und Lebensweise • Diversitätsmuster und Verbreitung in der Schweiz • Nicht-einheimische und invasive Arten • Faunistik der Amphipoden der Schweiz • Sammeln, Präparieren, Konservieren • Bestimmen	
Checkliste der Amphipoden der Schweiz	45
Bestimmungsschlüssel	49
Bestimmungsschlüssel der Familien und Gattungen	
Gattungsschlüssel	
Artsschlüssel	
Clés de détermination	104
Clé des genres	
Clé des espèces	
Identification keys	156
Key to genera	
Key to species	
Arttexte	209
Literatur	372
Index	387

156

Amphipoda

Key to genera

- 1 Accessory flagellum of antenna I absent (Plate 1.1). In some species, antenna I much smaller than antenna II (Plate 1.2). Telson fleshy, thick, short, or minute and usually uncleft. Mandibular palp vestigial or absent (dissection needed). Endopodit of uropod III always absent. → **Talitrida** and **Corophiida**

2

- Accessory flagellum of antenna I present (Plate 1.3). Antenna I and II of similar size or antenna I much longer than antenna II (Plate 1.4). Telson flat, laminar, movable and usually cleft, bilobed or at least distinctly incisive. Mandibular palp present (dissection needed). Endopodit of uropod III present in most species. → **Gammarida** and **Bogidiellida**

5

- 2 Antenna II enlarged and much stronger than gnathopods and pereopods (Plate 1.5). Pereopod VII longer or much longer than pereopod VI. Body dorso-ventrally flattened. Coxae I to IV small. Antenna II consisting of five segments without a distinct flagellum.

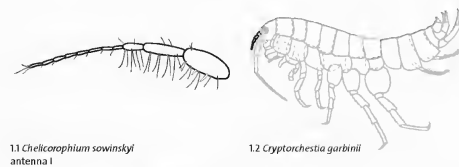
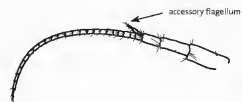
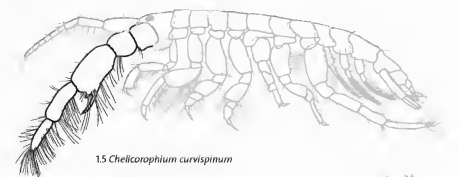
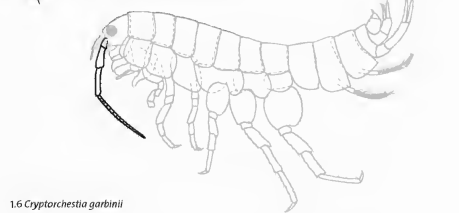
Chelicorophium

- Antenna II not enlarged (Plate 1.6). Pereopod VII of equal length as pereopod VI. Body laterally flattened. Coxae I to IV large. Antenna II with a distinct flagellum. → **Talitrida**

3

Key to genera

157

1.1 *Chelicorophium sowinskyi*
antenna I1.2 *Cryptorchestia garbinii*1.3 *Gammarus fossarum* s. l.
antenna I1.4 *Gammarus fossarum* s. l.1.5 *Chelicorophium curvispinum*1.6 *Cryptorchestia garbinii*

324

Amphipoda

Echinogammarus stammeri S. Karaman, 1931
(Tessiner Flohkrebs/italien-Flohkrebs)

Einzig einheimische Art der Gattung *Echinogammarus*. Sie kommt in der Schweiz nur im Tessin vor, wo sie in Bächen, Flüssen und Seen weit verbreitet und häufig ist. Dabei nimmt sie eine ähnliche ökologische Nische ein wie sie von *Gammarus fossarum* s. l. nördlich der Alpen belegt wird.

Typus Fundort

Monfalcone, Italien. Die aus der Schweiz beschriebene Art *Echinogammarus fluminensis* Pinkster & Stock, 1970 ist ein Synonym von *E. stammeri* (Karaman, 1993).

Allgemeine Verbreitung

Die Art kommt in Frankreich (Massiv de la Sainte Baume im Département Var), in Norditalien und auf dem Balkan bis nach Griechenland vor (Karaman, 1993). Sie ist vor allem in den Gewässern der Po-Ebene und deren Zuflüssen weit verbreitet.

Verbreitung in der Schweiz

Die Art kommt in der Schweiz nur im Tessin vor. Es handelt sich dabei um die nördliche Arealgrenze der norditalienischen Populationen. Sie ist vor allem in den tiefen Lagen des Tessins (Mendrisio, Lago di Lugano und Lago Maggiore und deren Zuflüsse) weit verbreitet.

Höhenverbreitung

Eine typische Art der tiefen Lagen. Von 200 m ü. M. (Lago Maggiore, Magadinoebene) bis 750 m ü. M. (höchster Nachweis in der Ticino bei Faido). Die meisten Vorkommen liegen unter 400 m ü. M.

Lebensraum

Echinogammarus stammeri ist eine Art mit einer sehr breiten ökologischen Nische. Sie kommt praktisch in allen aquatischen Lebensraumtypen in den tiefen Lagen des Tessins vor. Dies umfasst die grossen Seen (Lago Maggiore und Lago di Lugano), die grösseren Zuflüsse zu diesen Seen, aber auch die ganze Bandbreite an kleinen bis sehr kleinen Flüssen und Bächen bis hin zu Bewässerungskanälen im Landwirtschaftsgebiet (Abb. 96 & 97). Die Art kommt in schlammigem, sandigem bis grobschottrigem Substrat vor und nutzt auch die Makrophytenvegetation. Sie fehlt einzig in Gewässern, die entweder einer hohen Pestizidbelastung ausgesetzt sind oder die eine starke Schwall-Sunk-Dynamik aufweisen.

Im Tessin ist *E. stammeri* die am weitesten verbreitete Amphipodenart. An einigen Stellen kommen im gleichen Lebensraum auch *Synurella ambulans* respektive *Cryptorchestia garbinii* vor. Nach Stock (1968) kommt die Art in Italien auch zusammen mit *G. fossarum* s. l. und *G. pulex* vor.

Ökologie

Die Art nimmt im Tessin die ökologische Nische von *Gammarus fossarum* s. l. nördlich der Alpen ein. Sie ist an vielen Stellen sehr häufig und dürfte im aquatischen Nahrungsnetz

Arttexte

325

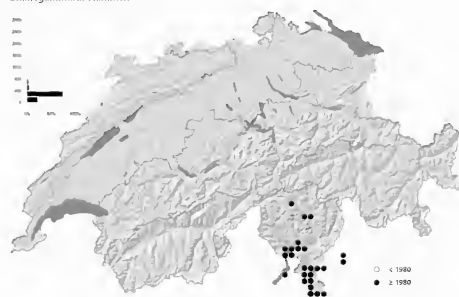
eine wichtige Funktion als Destruent respektive als Nährtier für Fische haben. Wie die Gammarus-Arten nördlich der Alpen kann *E. stammeri* durch das Auftreten neuer invasiver Arten massiv verdrängt werden. Casellaro et al. (2008) beschreiben dies anhand der Ausbreitung von *Dikerogammarus villosus* im Gardasee seit 2003.

Status

Einheimische Art.

Morphologie/Bestimmungsmerkmale

Habitus: Kleine bis mittelgrosse Amphipodenart (Körpergrösse um 12 mm). Die Art weist einen ähnlichen Habitus wie *Gammarus fossarum* s. l. auf, wobei sie etwas kleiner ist (Abb. 92). Sie ist durch ihre verkürzten Innenäste der Uropoden III einfach von der Gattung *Gammarus* zu unterscheiden. In Norditalien (Lombardien) und Frankreich (Einzugsgebiet Rhone) gibt es eine Reihe weiterer Arten der Gattung *Echinogammarus*, die möglicherweise bis in die Schweiz vorkommen und auf die bei der Bestimmung von *E. stammeri* geachtet werden muss. Ausführliche Bestimmungsschlüssel dieser Arten geben Karaman (1993) und Stock (1968). *Echinogammarus foxi* unterscheidet sich durch deutlich kürzere und robustere Pereopoden V-VII, wobei vor allem die Segmente 3-6 deutlich kürzer und weniger schlank sind als bei *E. stammeri*. Bei *Echinogammarus pungens*, *E. ruffoi* und *E. veneris* zeigt das Stielsegment 3 der Antenne I maximal die 2-2,5-fache Länge der Breite (bei *E. stammeri* ungefähr die dreifache Länge der Breite) und hat nur ein bis drei Gruppen ventraler Borsten. Bei *E. stammeri* hat dieses Segment vier bis fünf Gruppen ventraler Borsten (Abb. 98).

Echinogammarus stammeri

Patagonian amphipoda

I am writing from Puerto Madryn, Patagonia, Argentina, where several of us amphipodologists have attended the 10th International Conference on Marine Bioinvasion.

We were a little group of amphipod people from two continents:

Agnese Marchini from Italy, Macarena Ros & Gemma Martinez-Laiz from Spain, Anali' Campean from Perú, and Carlos Rumbold from Argentina.

All of us presented talks or posters with amphipod subjects, showing that this group has a relevant role in global Bioinvasions.

Anali' even presented the first ever survey of non-indigenous species in Perú, and amphipods resulted to be the dominant group!

This short report is just to update the amphipod newsletter on the Patagonian activities.

Best wishes from all of us!

Agnese Marchini

